Next generation sequencing in detecting Oral Cancer Due to Tobacco Consumption

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ABSTRACT: DNA sequence DNA Sequencing is the first step in establishing phylogenetic trees, protein structure prediction, diagnosis of cancer, discovery of drugs and hence its importance cannot be underestimated. DNA sequencing finds its use in the diagnosis of oral squamous cell carcinoma (OSCC). Oral Cancer is the most common occurring malignancies in the world, especially in India where the prevalence for smoking, Areca nut chewing coupled with a lifestyle that encourages these two activities as fashion are left many people diagnosed with OSCC. Patients with this OSCC are more likely unaware of its side effects and over time might suffer from facial deformity. The importance to understanding the symptoms, prevention and treatment of oral cancer is very much essential today. In this paper, we looked at over 2000 odd papers published and look at the correlation between the next Generation DNA sequencing algorithms (NGS) play an important role in diagnosis of OSCC. This is a further study on some of the papers which have highlighted NGS role in OSCC Diagnosis. We did like to see a comprehensive review on the papers published so far. In the discussion, we will see frequently mutated genes in the OSCC, recent discoveries and OSCC treatment based on the findings.

KEYWORDS: Sequence, phylogenetic Tree, Carcinoma, Gene.

1 INTRODUCTION

Tobacco is a consumed as a recreational substance either orally (chewed, sniffed, inhaled, and sucked) or through a third a party substance which contains tobacco, eg: Cigarette. Sometime, Tobacco is impersonalized as a cigarette in India when a tobacco is rolled like a small cigarette, eg Beedi [25,26]. In Latina America, thick tobacco leaves are fermented, and tobacco leaves of different grades are then layered to make a thick blunt which is popularly known as a cigar [27]. The consumption of tobacco increases the chances of oral cancer [11]. In India, although tobacco is still consumed orally, it is quickly fading away and is replaced by tobacco consumed through cigarette, cigar and beedis [27]. It is estimated that tobacco is the biggest killer, about 10 million deaths every year, of which tobacco consumption contributes to about 5 million [28,29]. From the literature, we observe that tobacco smokers have about 10 - 15% chance of developing oral cancer than nonsmokers and this percentage increases with the smoking frequencies [30-33].

Tobacco smoke and its byproducts mixed with saliva after chewing produces many carcinogenic materials which are commonly called polycyclic aromatic hydrocarbons (PAHs). These PAHs are more pronounced in smokers than orally consumed tobacco patients [34]. Apart from these PAHs, Nitrosamines, aromatic amines, and other agents which are present in elevated levels in a smoker's urine at any given point of time. The PAHs are the main causes for cellular transformation [35, 36]. These

PAHs would later alter, mutate the DNA, RNA, Protein and Genes in individuals. A RNA, DNA or a protein sequence is represented by a character set. DNA (A, C, G, T), RNA (A, C, G, U) and protein molecules (A, R, N, D, C, Q, E, G, H, I, L, K, M, F, P, S, T, W, Y, V) [1 - 9]. Tobacco smoking is quite addictive, and once addicted, there is now constant exposure to these carcinogenic substances. This exposure would now lead to bondage of carcinogens to DNA genome (molecular changes or DNA changes). This bondage would now lead to DNA mutation where a protein character at one place in entire genome is replaced by another protein, there by mutating into other DNA form. In some cases, the DNA character goes missing or a new protein is added when it should not be [34].

2   LITERATURE

Oral cancer can be defined as the presence of lesions or tumors in the lip or [10]. Oral Cancer is most common form of cancer seen across the world and its ranking within each continent varies from year to year. It occupies the highest ranking in India and surrounding south-east Asia [11, 12, 13]. From our literature review we could conclude that oral cancer diagnosis cases have more than doubled from 1990 - present. It's about 3 out of 100000 and the adjusted life expectancy is about 66 years from the data. In Southeast Asia, the prevalence and fashion driven cigarette smoking and areca nut chewing amounts, amounts to 98% of the cases. Some cases where the oral cancer is due to passive smoking, oral Hygiene and others are reported [15, 16].

Oral Squamous Cell Carcinoma (OSCC) is the most common malignant epithelial neoplasm affect the oral cancer. Oral cancer can affect any region including oral cavity, pharyngeal regions, and salivary glands. Most of the legions are prevalent in the anterior part of the tongue, Labial, gingiva, alveolar muscle, the palate and some cases in the parotid glands or salivary glands [11], [18 – 20]. Despite many technological advancements in the recent times both in detection, prognosis and treatment, common treatment for oral cancer is still surgery or radiotherapy. If the cancer is in its initial stage, then the survival rate is considerably higher with either surgery or radiotherapy [21]. But if the cancer is detected in the 3rd or 4th stage then the survival rate falls to 30% [21] and most of the patients die within 12 - 24 months if the cancer is the advanced stage [22 - 24].

Today, efforts are made to increase the efficacy of radiotherapy and chemotherapy, either through the advent of precision targeted therapy like laser, and targeted medicine to expedite the research and development of targeted therapies and screening strategies in the oral cancer [18], [19]. Targeted molecular therapy, like monoclonal antibodies and gene therapy, have been applied to oral cancer patients. These molecules are associated with the proliferation of OSCC [20].

These protein displacement or replacement in the DNA chain play the pivotal role in the advert mutation and progression of the OC. DNA genome project is perhaps the biggest project ever taken in the bioinformatics area in the early 1990s [5]. Ever since then, the appetite for DNA knowledge acquisition has grown tremendously. With this curiosity, the research produced DNA sequencing algorithms ranging from local sequence alignment algorithms such as Smith-Waterman [37], FASTA [38], BLAST [39], GappedBLAST [40], BLASTZ [41], PatternHunter [42], YASS [43], LAMBDA [44], USearch [45], LAST [46], and ALLAlign [47] and MASAA [1-4]. These algorithms are concentrating on aligning locally similar sequences.

There are other algorithms which concentrate on aligning the entire sequences (Pairwise), these are Needleman and Wunsch [48], GLASS [49], WABA [50], and GLASS [51]. MASAA, GLASS and BLASTZ are very important in this research, in that, these algorithms a novel method of first finding regions of similarity quickly and then employ another novelty called stitching algorithm to finally find the final alignment. The regions of similarity size, and the stitching algorithm vary in these algorithms, which brings to novelty surge in multiple sequence alignment. When the need to align multiple sequences, where more than 2 sequences are aligned to find regions, came about, more researchers now had to look at the above algorithms to find regions of similarity quickly yet wanted alignment that made biological sense.

The popular multiple sequence alignment algorithms were ClustalW [52], MAFFT [53], Kalign [54], Probalign [55], MUSCLE [56], DIALIGN [57], PRANK [58], FSA [59], T – Coffee [60, 61], and Probcons [62]. Other approaches focused on Genetic and motifs algorithms [5]. Genetic algorithms got traction, while motifs-based algorithm was not successful [5].

3   NEXT GENERATION SEQUENCING

Next-generation sequencing (NGS) is a technique where powerful massive computer clusters are run to parallelly sequence millions of small segments of DNA. NGS enables researchers to study genomes amongst many sequence types systematically and make insights which in turn leading into disease understanding and drug discovery. In NGS, the entire genome or targeted segments are sequenced quickly [10]. As discussed in the previous section, researchers were prodding new algorithms ever since DNA sequence are discovered, for more insights [5]. Frederick Sanger in 1970s popularised a sequencing method using a 'plus and minus' method. It involved the usage of phix174 and chain-terminating inhibitors [11, 12]. There was Maxam-Gilbert

which gained popularity for a brief period and quickly the method was abandoned. as this method was cumbersome and involved the use of many chemicals [14]. Sanger method is still considered to be one of the classical methods and is still in use today.

NGS is used in two platforms Illumina/Solexa and Ion Torrent. Of these two, the Illumina/Solexa is a low cost and high yield method, which has propelled it to be the preferred platform today. The Ion Torrent, method is rather quick and produces longer read lengths than the Illumina [15], however due to the method being expensive, it is not used widely today [14, 15]. Other platforms like the Oxford Nanopore Technologies (ONT) and Pacific Biosciences (PacBio) are quite new and they too produce longer lengths, however, from the literature, the price to use is not yet clear [16]. Asia especially China has poured enormous amount of money in developing new platforms, one of method coming out of ASIA is Beijing Genomic Institutes BGISEQ, standing for Beijing Genomic Institute Sequencing. BGISEQ used a new nanotechnology to generate terabytes of sequenced data [63-65]. This method is also less expensive but produces longer length compared to Illumina. It is to be seen in the future if this method would be adopted widely as illumina/Solexa is today [64,66].

One of the fields that has immensely benefitted from NGS is microbiology and dentistry. NGS has played an important role in identifying oral microbiota in humans. These newly identified microorganisms which live in human oral cavity is thus enlarging the dentistry knowledge base to support good overall good health. Two methods are used to evaluate the kind of organisms present in the oral cavity using NGS is called 16 rRNA sequencing and the other shotgun sequencing [67]. 16 rRNA is present in all bacteria and therefore this knowledge can be used to know whether the bacteria are present in one's oral cavity or not [68]. Shot sequencing is used to gain knowledge on microbes' genome and resistance of genes to certain conditions [69]. Using these two methods, NGS can now classify different microbial genomes, this knowledge is now used for disease diagnosis, germ or bacterial resistance know-how and drug discovery.

NGS has overcome some of the limitations of PCR based sequencing, Sanger sequencing, and other micro array and SNP array methods, either through better sensitivity or better genomic coverage or both. This better sensitivity using NGS has through more light on some of the ambiguities in oral cancer classification. In other words, Sanger sequencing uses a minimum mutant percentage of about 25%, this is quite large, in fact, some cancer could now be diagnosed well below 25% of mutant allele [70,71]. The PCR method is quite limited to only few know sequences in detecting the tumours or oral cancer [72]. NGS expands this horizon as it can sequence the entire genome, targeted sequencing, RNA, and mRNA too [73]. Today, research has shown the tumour after biopsies are heterogenous in nature, this now presents a difficult proposition if PCR method is used, as it would not be able to detect the mutation (gene) less of 25%. NGS would otherwise be able to detect such kind of solid heterogenous tumours [74,75]. In this paper, we will review what are the genes which get modified in patients with smoking habits resulting in oral cancer using NGS technology. We do this by going through some 1800 plus papers who used NGS technology and what genes their research show cased, and we draw our conclusion from this exhaustive research.

4   RESEARCH METHODOLOGY

Our literature search was conducted according to PRISMA guidelines, where in all studies which employed NGS technology to study the impact of smoking and its direct correlation to oral cancer was studied [77-79]. We expanded the already existing study made by [76] to be broader and more exhaustive as the previous study conducted sample size was skewed towards records which did not have any NGS technology and only a 10% out of 1200 odd papers had any mention of Oral cancer and NGS simultaneously. We felt this was not exhaustive study and hence brought upon ourselves to expand appropriately to reflect the real picture of the NGS and Oral cancer correlation. We searched through freewheeling, along with Pubmed and Embase records and we want to emphasize that new expanded records all have NSG and Oral Cancer. As we searched, many duplicates, derived research papers were removed, then a consensus number were taken into the final pool was finalized.

5   METHOD

The Records were researched from Jan 2023 till September 2023 on PubMed, Embase, Researchgate and ScienceDirect and freewheeling. A total of 1921 records were researched, out of which any mention of NGS and oral cancer was 1505 records. The authenticity of the records was examined by 2 independent reviewers. Both primary and secondary and data extraction, from the papers were done by both the researchers. There were approximately 200 records bring the combined examined papers to 2021 papers. 1: There were records on Basic Analysis of NSG, Smoking and Oral Cancer (n = 29), 2: records on Diagnosis based and Analysis (n = 1), 3: records on just model (n = 1), 4: records on NSG, Smoking and oral cancer (n = 33), 5: Records on NGS but no data (n =3), 6: Records not on either NGS or Oral Cancer or Smoking or any combination of these 2 (n = 127), 7: Records on reviews (n = 52), 8: Record not on oral cancer (n = 1249).
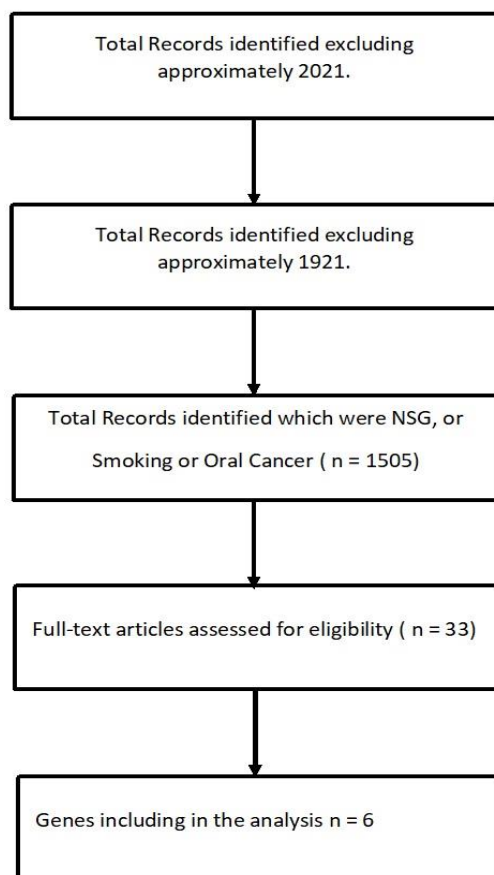
Fig. 1. Flow chart of workflow

Oral cancer or OSCC or oral or squamous cell carcinoma and next generation sequencing or NGS or Smoking or DNA sequencing and Genes was used in Embase and fields like ("oral") OR ("Carcinom") OR ("Oral Cancer") or ("Mouth Cancer") Or Its combinations OR ("carcinoma" [All Fields] AND "squamous" [All Fields] AND "cell" [All Fields]) OR "squamous cell carcinoma" [All Fields] OR ("squamous" [All Fields] AND "cell" [All Fields] AND "carcinoma" [All Fields])) AND ("mouth" [MeSH Terms] OR "mouth" [All Fields] OR AND "cavity" [All Fields]))) OR (("carcinoma, squamous cell" [MeSH Terms] OR ("carcinoma" [All Fields] AND "squamous" [All Fields] AND "cell" [All Fields]) OR "squamous cell carcinoma" [All Fields] OR ("squamous" [All Fields] AND "carcinoma" [All Fields])) OR "tongue" [All Fields])) was used in pubmed and oral cancer, smoking on freewheeling [76].Records that utilized NGS, Smoking, and genes described in OC patients with smoking were 33 with genes considered majorly is 6. The entire workflow is shown in Figure 1.

6   RESULTS AND ANALYSIS

There were 33 included in this research, with full text with 34 genes found in OC patients who smoke (Figure 2 and Table S1). Of these there are 6 genes which were found in more than 4 records. These were 11p15.5 (HRAS), 17p13.1 (TP53), 9p21.3 (CDKN2A), 9q34.3 (NOTCH1), and 4q35.2 (FAT1) and 4q31.3 (FBXW7) and 9p24.1 (PD-L1) were found to be in 3 records and the rest of the genes were found to be 1 record. This is represented in the figure 2.
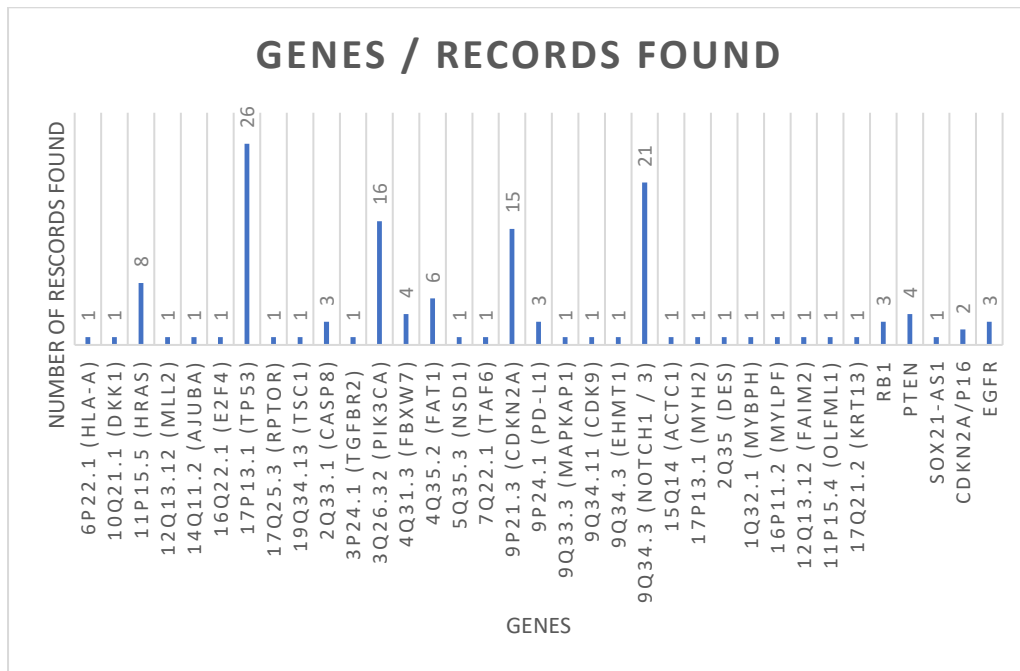
Fig. 2.   Records examined vs genes found in record

From the literature, we can confidently say that smoking is directly correlated with oral cancer and the chances for OC diagnosis raises if the patient smoking frequency of smoking is high (chain smokers). It is also found out the smoking somehow is responsible for genome mutation, transcriptomic changes, some of these changes are found in some genes portion of the genome, which has catastrophic effect on the changes inside the cell (cell reaction and protein developed, and expressions). All these changes are quite complex and leads to the initiation of the oral cancer [118]. Tp53 is a gene responsible for cell health and its reproduction (growth) [118, 119]. Since it is a important gene in cell health, it also plays a pivotal role in suppressing any changes to the cell health, structure or multiplication [118, 119]. Alternation to this gene could be seen, as the cancer origination and spread of cancer [119]. From the literature, we can say with certain surety that mutations in the Tp53 gene were present in various human cancer [118]. However, when it comes to oral cancer, we have found from our research, that mutation in Tp53 were found in 78% of the 33 records, which is 16 in number [80 - 91]. This is quite an observation, we have found various authors [81 - 89, 91] and [118], believe and observe that this gene mutation is the primary initiator for the Oral Cancer proliferation. These authors [81 - 89, 91] also say that missense mutations are the most common. It is also observed that Tp53 is located near DNA binding are of the protein [81 - 89, 91] and it is associated with insufficient or poor prognosis [81 - 85]. Authors in [120] have noticed that there is common change or mutation is from G -> A or G -> T, and the authors observe that this mutation scenarios have more to do with smoking and consumption of tobacco in one form or the other. Authors in [121] have observed that such mutations have increases spread of OC, increased penetration, and increased resistance to drugs and eventually leading to death.

NOTCH1 is another gene mutated in OC associated with smoking. NOTCH1 is also a tumour suppressor gene very similar to TP53 and similarly to Tp53, any changes to this gene leads to initiation and multiplication of cancerous cells (tumour) [80 - 91]. From our research, we found out that, 63% of the records analysed or full text record had NOTCH1 gene mutated, which is a substantial percentage than the percentage found on other review [76]. We also found out from the records we looked at; all these studies were predominantly Asians (Including South - Asia). This tells us that NOTCH1 mutation is a likely event in all OC cancerous patients [118] in samples from Asia. Authors [118] evident that this observation is predominantly because of the lifestyle of population, with many consuming locally made cigarettes (Bedis in India, Secondary smoking (smoking from discarded cigarettes)) and alcohol consumption. In our research, we saw one NOTCH gene variation and this NOTCH variation was a study made in Europe, and we are now certain that this variation is also in other races but perhaps not in scale or volume found in Asian population. The mutation in NOTCH1 genes were found in different location all along the length of the exon [118, 119] and such mutations in all studies clearly point to decreased confidence in the survival of the patient [91].

The third gene which was found to be highly mutated in records was PIK3CA, this gene accounted for 48% which is quite less than the other 2 genes found above. The exon 20 is the common area mutated of PIK3CA [86-89] and this affected area has been seen to be associated with lower lip region of the patient [86]. Authors of [82] have observed that mutation in the

PIK3CA has a strong correlation with the later stage of OC, clearly showing that this mutation is clear signal that patient prognosis would be bad when this gene is mutated. Hence, a different targeted therapy to rectify this situation is highly recommended in the literature. Authors in [83] have observed that the activation of PI, suggest that Signalling pathway could be improved either by targeted treatment of other means. From the research, we conclude that a mutation of PIK3CA is a clear signal that OC is the later stage, prognosis could be hampered, a clear sign that alternative therapy should be looked and provided to the patient immediately, if not, then the patient is looking at grim and sad prognosis leading to fatality.

CDKN2A gene is associated with cell cycle inhibitor [122]. Authors have found that a downregulated expression of this gene in the life cycle of the cell correlates with OC and suggest that this gene could now be used a marker for OD diagnosis. In our research, we found 47% of the full text records to have this gene mutated with the OC occurrence and unlike the other genes, we could not correlate this gene mutation with any region of the world [123]. Most of the reported mutation in the records suggest that there is mutation from G -> T protein in the exon 2 and in position 322 of this exon, and this correlated with primary head and neck cancers along with oral cavity [124]. This gene mutation is also correlated with OC recurrence after the OC was partially cured in the patient, meaning, that the cell cycle inhibitor played an important role in recurrence of OC [125]. We also observed that majority of the tongue related OC had CDKN2A gene mutated. The prognosis was poor after this CDKN2A was mutated, leading the poor prognosis, indicating the current therapy must be stopped and alternative therapy be initiated to curb OC growth [126].

H-RAS gene is also an important regulate cell health, differentiation, migration, and apoptosis [80]. Mutation of this associated with occurrence of OC [80 - 91]. Incidentally, HRAS genes is quite less often seen in OC than other cancers [121 - 125], however, in neck and head and oral cancers, we observe that this gene is seen [80 - 91], [119 - 123, 125]. In our study we found H-RAS gene studies to be around 25%, this gene is also associated from the research to be more prevalent in patients who chew tobacco and areca nuts with alcohol consumption. From our study, we conclude that H-RAS gene to be indicator of OC in the later stage and an observation of this gene mutation correlated to patient living with poor condition, consumption of alcohol, smoking, tobacco chewing or areca nut chewing along with betel leaves.

FAT-1 gene role in cancer has been controversial in nature. FAT1 acts as a cancer suppression role especially in HNSC and oral cancers. The expression of this gene is mostly seen in patients with later stages of OC. In our study, we saw 18% of the records mentioning FAT1 gene [83]. We observed from the records, any mention of FAT1, was seen the stage 4 compared to other cancers [89 – 91]. Also, the samples included in the research records showed that OC was lip and oral cavity (Roof) than other regions in OC. FAT1 gene was also not associated with any world region or sample lifestyle. Prognosis was observed to be poor leading to fatality in 2 of the records.

7   CONCLUSION

In this study, we included 33 records with full article reads on OC, NGS and smoking correlation. We found some interesting observation, several of them, revealed that certain gene is mutated in some ethnicities than the rest, genes were mutated less than others in various regions. The correlation of certain gene mutation to lifestyle was also observed. The study has revealed that we need more research on OC to accurately diagnose and treat early-stage OC. This study was also on the NGS sequencing technique used to find OC with samples or records on smoking or areca nut consumption. This study revealed certain gene, most of the times, the first 2 genes mentioned in the study to be mutated as these are OC inhibitors. Smoking is a killer, and it is incumbent on all people in all regions around of the world to educate people on repercussions of tobacco addiction either through smoking or chewing.

APPENDIX

All data on the records is in the excel file ResearchTableWithTabs_S1S2S3.

CONFLICT OF INTEREST

The authors have no conflicts of interest in this research.

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REFERENCES

[1] Reddy, Bharath Govinda. *Multiple Anchor Staged Local Sequence Alignment Algorithm-MASAA*. Diss. The University of Northern British Columbia, 2009.

[2] Waqar Haque, Alex Aravind, and Bharath Reddy. 2009. Pairwise sequence alignment algorithms: a survey. In Proceedings of the 2009 conference on Information Science, Technology and Applications (ISTA '09). ACM, New York, NY, USA, 96-103. DOI=http://dx.doi.org/10.1145/1551950.1551980.

[3] Haque, W., Aravind, A., & Reddy, B. (2008, August). An efficient algorithm for local sequence alignment. In *2008 30th annual international conference of the IEEE engineering in medicine and biology society* (pp. 1367-1372). IEEE.

[4] B. Reddy and R. Fields, «Multiple Anchor Staged Alignment Algorithm – Sensitive (MASAA – S),» *2020 3rd International Conference on Information and Computer Technologies (ICICT),* San Jose, CA, USA, 2020, pp. 361-365, doi: 10.1109/ICICT50521.2020.00064.

[5] Reddy, B., Fields, R. (2022). Multiple Sequence Alignment Algorithms in Bioinformatics. In: Zhang, YD., Senjyu, T., So-In, C., Joshi, A. (eds) Smart Trends in Computing and Communications. Lecture Notes in Networks and Systems, vol 286. Springer, Singapore. https://doi.org/10.1007/978-981-16-4016-2_9.

[6] Bharath Reddy and Richard Fields. 2022. From past to present: a comprehensive technical review of rule-based expert systems from 1980 -- 2021. In Proceedings of the 2022 ACM Southeast Conference (ACM SE '22). Association for Computing Machinery, New York, NY, USA, 167–172. https://doi.org/10.1145/3476883.3520211.

[7] Reddy, B., & Fields, R. (2023). Maximum Match Subsequence Alignment Algorithm Finely Grained (MMSAA FG). *arXiv e-prints,* arXiv-2305.

[8] Reddy, M. (2020). BIOINFORMATICS & PAIRWISE SEQUENCE ALIGNMENT: Local and Global Sequence Alignment Algorithms. Independently published.

[9] Bharath Reddy and Richard Fields, «Techniques for Reader-Writer Lock Synchronization,» International Journal of Electronics and Electrical Engineering, Vol. 8, No. 4, pp. 63-73, December 2020. doi: 10.18178/ijeee.8.4.63-73.

[10] Behjati, S.; Tarpey, P.S. What is next generation sequencing? Archives of disease in childhood. Education and practice edition. BMJ J. 2013, 98, 236–238.

[11] Sanger, F.; Air, G.M.; Barrell, B.G.; Brown, N.L.; Coulson, A.R.; Fiddes, J.C.; Hutchison, C.A.; Slocombe, P.M., Smith, M.; Sanger, G.M.A.F. Nucleotide sequence of bacteriophage 'X174 DNA. Nature 1977, 265, 687–695.

[12] Sanger, F.; Nicklen, S.; Coulson, A.R. DNA sequencing with chain-terminating inhibitors. Proc. Natl. Acad.Sci. USA 1977, 74, 5463–5467.

[13] Maxam, A.M.; Gilbert, W. A new method for sequencing DNA. Proc. Natl. Acad. Sci. USA 1977, 74, 560–564.

[14] Hodzic, J.; Gurbeta, L.; Omanovic-Miklicanin, E.; Badnjevic, A. Overview of Next-generation Sequencing Platforms Used in Published Draft Plant Genomes in Light of Genotypization of Immortelle Plant (Helichrysiumarenarium). Med. Arch. 2017, 71, 288–292.

[15] Liu, L.; Li, Y.; Li, S.; Hu, N.; He, Y.; Pong, R.; Lin, D.; Lu, L.; Law, M. Comparison of Next-Generation Sequencing Systems. J. Biomed. Biotechnol. 2012, 2012, 1–11.

[16] Oxford Nanopore bests PacBio. Nat. Biotechnol. 2019, 37, 336.

[17] Choi S, Myers JN. Molecular pathogenesis of oral squamous cell carcinoma: implications for therapy. J Dent Res. 2008; 87: 14–32.

[18] Loomans-Kropp, H.A.; Umar, A. Cancer prevention and screening: The next step in the era of precision medicine. NPJ Precis. Oncol. 2019, 3, 3.

[19] Price, C.P. Point of care testing. BMJ 2001, 322, 1285–1288.

[20] Hamakawa H, Nakashiro K, Sumida T, et al. Basic evidence of molecular targeted therapy for oral cancer and salivary gland cancer. Head Neck. 2008; 30: 800–9.

[21] Marsh D, Suchak K, Moutasim KA, et al. Stromal features are predictive of disease mortality in oral cancer patients. J Pathol. 2011; 223: 470–81.

[22] de Araújo RF, Barboza Jr CA, Clebis NK, de Moura SA, Lopes Costa Ade L. Prognostic significance of the anatomical location and TNM clinical classification in oral squamous cell carcinoma. Med Oral Pathol Oral Cir Bucal. 2008; 13: E344–7.

[23] Hill BT, Price LA. Lack of survival advantage in patients with advanced squamous cell carcinomas of the oral cavity receiving neoadjuvant chemotherapy prior to local therapy, despite achieving an initial high clinical complete remission rate. Am J Clin Oncol. 1994; 17: 1–5.

[24] Zini A, Czerninski R, Sgan-Cohen HD. Oral cancer over four decades: epidemiology, trends, histology, and survival by anatomical sites. J Oral Pathol Med. 2010; 39: 299–305.

[25] Digard, Helena, Proctor Christopher, Kulasekaran Anuradha, Malmqvist Ulf, and Richter Audrey. 2013. «Determination of Nicotine Absorption From Multiple Tobacco Products and Nicotine Gum.» Nicotine & Tobacco Research 15 (1): 255–61.https://doi.org/10.1093/ntr/nts123.

[26] Ansari, Zaki Anwar, S. Nafees Bano, and M. Zulkifle. 2010. «Prevalence of Tobacco Use Among Power Loom Workers—A Cross Sectional Study.» Indian Journal of Community Medicine 35 (1): 34–39. https://doi.org/10.4103/0970-0218.62551.

[27] Rassool, Hussein G. 2004. «Current Issues and Forthcoming Events.» Journal of Advanced Nursing 46 (3): 337–41. https://doi.org/10.1111/j.1365-2648.2004.02995.x.

[28] WHO Tobacco Fact Sheet. 2020. https://www.who.int/newsroom/fact-sheets/detail/tobacco.

[29] The Tobacco Connection. n.d. https://oralcancerfoundation.org/understanding/tobacco/.

[30] Stransky, Nicolas, Ann M. Egloff, Aaron D. Tward, Aleksandar D. Kostic, Kristian Cibulskis, Andrey Sivachenko, and Gregory V. Kryukov, et al. 2011. «The Mutational Landscapeof Head and Neck Squamous Cell Carcinoma.» Science333 (6046): 1157–60. https://doi.org/10.1126/science.1208130.

[31] Agrawal, Nishant, J. Frederick Mitchell, Curtis R. Pickering, Chetan Bettegowda, Kyle Chang, Ryan J. Li, Carole Fakhry, et al. 2011. «Exome Sequencing of Head and Neck Squamous Cell Carcinoma Reveals Inactivating Mutations in NOTCH1.» Science 333 (6046): 1154–57. https://doi.org/10.1126/science.1206923.

[32] Pickering, Curtis R., Jiexin Zhang, David M. Neskey, Mei Zhao, Samar A. Jasser, Jiping Wang, Alexandra Ward, et al. 2014. «Squamous Cell Carcinoma of the Oral Tongue in Young Non Smokers Is Genomically Similar to Tumors in Older Smokers.» Clinical Cancer Research 20 (14): 3842–48.https://doi.org/10.1158/1078-0432.CCR-14-0565.

[33] Upadhyay, Pawan, Nair Sudhir, Kaur Ekjot, Aich Jyotirmoi, Dani Prachi, Gardi Nilesh, and Sethunath Vidyalakshmi, et al. 2016. «Notch Pathway Activation Is Essential for Maintenance of Stem-Like Cells in Early Tongue Cancer.» Oncotarget 7 (31): 50437–49. https://doi.org/10.18632/oncotarget.10419.

[34] How Tobacco Smoke Causes Disease. 2010. The Biology and Behavioural Basis for Smoking-Attributable Disease: A Report of the Surgeon General. Centres for Disease Control and Prevention (US).

[35] Nagaraj, Nagathihalli S., Simone Beckers, John K. Mensah, Sabine Waigel, Nadarajah Vigneswaran, and Wolfgang Zacharias. 2006. «Cigarette Smoke Condensate Induces Cytochromes P450 and Aldo-Keto Reductases in Oral Cancer Cells.» Toxicology Letters 165 (2): 182–94. https://doi.org/10.1016/j.toxlet.2006.03.008

[36] Mahadevan, Brinda, Keshava Channa, Musafia Jeknic Tamara, Pecaj Arta, Weston Ainsley, and Baird William M. 2005. «Altered Gene Expression Patterns in MCF 7 CellsInduced by the Urban Dust Particulate Complex Mixture Standard Reference Material 1649a.» Cancer Research 65 (4): 1251–58. https://doi.org/10.1158/0008-5472.CAN-04-2357.

[37] Smith, T.F. and Waterman, M.S. 1981. Identification of common molecular subsequences. J. Mol. Biol. 147: 195-197.

[38] D. Lipman and W. Pearson. Rapid and sensitive protein similarity searches. Science, 227: 1435, 1441, 1985.

[39] Altschul, S.F., Gish, W., Miller, W., Myers, E. W., and Lipman, D. J. 1990. Basic local alignment search tool. J. Mol. Biol. 215: 403-410.

[40] S. F. Altschul, T. L. Madden, A. A. Schaer, J. Zhang, Z. Zhang, W. Miller, and D. J. Lipman. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Research, 25 (17): 3389, 3392, 1997.

[41] Schwartz, S., Zhang, Z., Frazer, K.A., Smit, A., Riemer, C., Bouck, J., Gibbs, R., Hardison, R., and Miller, W. 2000. PipMaker-A web server for aligning two genomic DNA sequences. Genome Res. 10: 577-586.

[42] Ma, B., J. Tromp, and M. Li (2002). Patternhunter: Faster and more sensitive homology search. Bioinformatics 18, 440-445.

[43] Laurent No and Gregory Kucherov. YASS: enhancing the sensitivity of DNA similarity search. Nucleic Acids Res, 2005.

[44] Hauswedell, H., Singer, J. & Reinert, K. Lambda: the local aligner for massive biological data. Bioinformatics 30, i349–i355 (2014).

[45] Edgar (2010) Edgar RC. Search and clustering orders of magnitude faster than BLAST. Bioinformatics. 2010; 26 (19): 2460–2461. doi: 10.1093/bioinformatics/btq461.

[46] Adaptive seeds tame genomic sequence comparison. Kiełbasa SM, Wan R, Sato K, Horton P, Frith MC. Genome Res. 2011 21 (3): 487-93.

[47] Wachtel. E. All ALL Local Alignment. http://www.allalign.com/.

[48] Needleman, S.B. and Wunsch, C.D. 1970. A general method applicable to the search for similarities in the amino acid sequence of two proteins. J. Mol. Biol. 48: 443-453.

[49] Delcher A.L., Kasif S., Fleischmann R.D., Peterson J., White O., and Salzberg S.L. 1999. Alignment of whole genomes. Nucleic Acids Res. 27: 2369-2376.

[50] Kent, J. and Zahler, M. 2000. The Intronerator: Exploring introns and alternative splicing in C. elegans genomic alignment. Genome Res. 10: 1115-1125.

[51] Bray Nick, Dubchak Inna and Pachter Lior, Avid: A global alignment program, Genome Research. 2003 13: 97-102; 2002.

[52] J. D. Thompson, D. G. Higgins, and T. J. Gibson, «CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice,»Nucleic Acids Research, vol. 22, no.22, pp.4673–4680,1994.

[53] K. Katoh and D. M. Standley, «MAFFT multiple sequence alignment software version 7: improvements in performance and usability,» Molecular Biology and Evolution, vol. 30, no. 4, pp.772–780,2013.

[54] T. Lassmann and E. L. L. Sonnhammer, «Kalign—an accurate and fast multiple sequence alignment algorithm,» BMC Bioinformatics, vol.6, article298,2005.

[55] U. Roshan and D. R. Livesay, «Probalign: multiple sequence alignment using partition function posterior probabilities,» Bioinformatics, vol.22, no.22, pp.2715–2721,2006.

[56] R.C.Edgar, «MUSCLE: a multiple sequence alignment method with reduced time and space complexity,» BMCBioinformatics, vol.5, article113,2004.

[57] B. Morgenstern, «DIALIGN: multiple DNA and protein sequence alignment at BiBiServ,» Nucleic Acids Research, vol.32, supplement2, pp. W33–W36,2004.

[58] A. L¨oytynoja and N. Goldman, «Phylogeny-aware gap placement prevents errors in sequence alignment and evolutionary analysis,» Science, vol.320, no.5883, pp.1632–1635,2008.

[59] R. K. Bradley, A. Roberts, M. Smoot et al., «Fast statistical alignment,» PLoS Computational Biology, vol. 5, no. 5, Article IDe1000392,2009.

[60] P. Di Tommaso, S. Moretti, I. Xenarios et al., «T-Coffee: a webserver for the multiple sequence alignment of protein and RNA sequences using structural information and homology extension,» Nucleic Acids Research, vol. 39, supplement 2, pp. W13–W17,2011.

[61] C. Notredame, D.G. Higgins, and J. Heringa, «T-coffee: a novel method for fast and accurate multiple sequence alignment,» JournalofMolecularBiology, vol.302, no.1, pp.205–217,2000.

[62] C.B.Do, M.S.P. Mahabhashyam, M. Brudno, and S. Batzoglou, «ProbCons: probabilistic consistency-based multiple sequence alignment,» GenomeResearch, vol.15, no.2, pp.330–340,2005.

[63] Li, Q.; Xia, Z.; Zhang, W.; Wang, L.; Wang, J.; Xu, D.; Mei, Z.; Liu, Q.; Du, S.; Li, Z.; et al. Reliable multiplex sequencing with rare index mis-assignment on DNB-based NGS platform. BMC Genom. 2019, 20, 215.

[64] Zhu, F.-Y.; Chen, M.-X.; Ye, N.-H.; Qiao, W.-M.; Gao, B.; Law, W.-K.; Tian, Y.; Zhang, N.; Zhang, D.; Liu, T.-Y.; et al. Comparative performance of the BGISEQ-500 and Illumina HiSeq4000 sequencing platforms for transcriptome analysis in plants. Plant Methods 2018, 14, 69.

[65] Huang, J.; Liang, X.; Xuan, Y.; Geng, C.; Li, Y.; Lu, H.; Qu, S.; Mei, X.; Chen, H.; Yu, T.; et al. A reference human genome dataset of the BGISEQ-500 sequencer. GigaScience 2017, 6, 1–9.

[66] Mak, S.S.T.; Gopalakrishnan, S.; Carøe, C.; Geng, C.; Liu, S.; Sinding, M.H.S.; Kuderna, L.; Zhang, W.; Fu, S.; Vieira, F.G.; et al. Comparative performance of the BGISEQ-500 vs Illumina HiSeq2500 sequencing platforms for palaeogenomic sequencing. GigaScience 2017, 6, 1–13.

[67] Zaura, E. Next-generation Sequencing Approaches to Understanding the Oral Microbiome. Adv. Dent. Res. 2012, 24, 81–85.

[68] Kilian, M.; Chapple, I.L.C.; Hannig, M.; Marsh, P.D.; Meuric, V.; Pedersen, A.M.L.; Tonetti, N.S.; Wade, W.G.; Zaura, E. The oral microbiome—An update for oral healthcare professionals. Br. Dent. J. 2016, 221, 657–666.

[69] Deurenberg, R.H.; Bathoorn, E.; Chlebowicz, M.A.; Couto, N.; Ferdous, M.; García-Cobos, S.; Kooistra-Smid, A.M.; Raangs, E.C.; Rosema, S.; Veloo, A.C.; et al. Application of next generation sequencing in clinical microbiology and infection prevention. J. Biotechnol. 2017, 243, 16–24.

[70] Zhao, Y. A comparative study of ddPCR and sanger sequencing for quantitative detection of low-frequency mutation rate. IOP Conf. Ser. Earth Environ. Sci. 2019, 332, 032023.

[71] Lin, M.-T.; Mosier, S.L.; Thiess, M.; Beierl, K.F.; Debeljak, M.; Tseng, L.-H.; Chen, G.; Yegnasubramanian, S.; Ho, H.; Cope, L.; et al. Clinical Validation of KRAS, BRAF, and EGFR Mutation Detection Using Next-Generation Sequencing. Am. J. Clin. Pathol. 2014, 141, 856–866.

[72] Smith, C.J.; Osborn, A.M. Advantages, and limitations of quantitative PCR (Q-PCR) -based approaches in microbial ecology. FEMS Microbiol. Ecol. 2009, 67, 6–20.

[73] Azim, F.S.; Houri, H.; Ghalavand, Z.; Nikmanesh, B. Next Generation Sequencing in Clinical Oncology: Applications, Challenges and Promises: A Review Article. Iran. J. Public Health 2018, 47, 1453–1457.

[74] Gabusi, A.; Gissi, D.B.; Tarsitano, A.; Asioli, S.; Marchetti, C.; Montebugnoli, L.; Foschini, M.P.; Morandi, L. Intratumoral Heterogeneity in Recurrent Metastatic Squamous Cell Carcinoma of the Oral Cavity: New Perspectives A orded by Multiregion DNA Sequencing and mtDNA Analysis. J. Oral Maxillofac. Surg. 2020,77, 440–455.

[75] Gabusi, A.; Gissi, D.B.; Montebugnoli, L.; Asioli, S.; Tarsitano, A.; Marchetti, C.; Balbi, T.; Helliwell, T.R.; Foschini, M.P.; Morandi, L. Prognostic impact of intra-field heterogeneity in oral squamous cell carcinoma. Virchows Arch. 2019, 476, 585–595.

[76] Sharma, Vishwas & Kumar, Dinesh & Kumar, Shravan & Singh, Harpreet & Sharma, Naveen & Gupta, Sanjay. (2022). Impact of tobacco smoking on oral cancer genetics—A next-generation sequencing perspective. iMeta. 1. 10.1002/imt2.44.

[77] Liberati, Alessandro, Douglas G. Altman, Jennifer Tetzlaff, Cynthia Mulrow, Peter C. Gøtzsche, John P. A. Ioannidis, Mike Clarke, et al. 2009. «The PRISMA Statement for Reporting Systematic Reviews and Meta-Analyses of Studies That Evaluate Health Care Interventions: Explanation and Elaboration.» Journal of Clinical Epidemiology 62 (10): e1–34. https://doi.org/10.1016/j.jclinepi.2009.06.006

[78] Gao, Haihe, Lisha Li, Mang Xiao, Yongwei Guo, Yi Shen, Lixin Cheng, and Ming Tang. 2018. «Elevated DKK1 Expression Is an Independent Unfavorable Prognostic Indicator of Survival in Head and Neck Squamous Cell Carcinoma.» Cancer Management and Research 10: 5083–89. https://doi.org/10.2147/CMAR.S177043.

[79] Zain, Rosnah B., L. P. Karen, Sok Ching Cheong, Arif Anwar, wan mahadzir Mustaffa, N. Prepagaran, Zuraiza Mohamad Zaini, et al. 2011. «Effects of Smoking on Oral Cancer Transcriptome.» Oral Oncology 47: S48. https://doi.org/10.1016/j.oraloncology.2011.06.169

[80] Song, X.; Xia, R.; Li, J.; Long, Z.; Ren, H.; Chen, W.; Mao, L. Common and complex Notch1 mutations in Chinese oral squamous cell carcinoma. Clin. Cancer Res. 2014, 20, 701–710.

[81] Nakagaki, T.; Tamura, M.; Kobashi, K.; Omori, A.; Koyama, R.; Idogawa, M.; Ogi, K.; Hiratsuka, H.; Tokino, T.; Sasaki, Y. Targeted next-generation sequencing of 50 cancer-related genes in Japanese patients with oral squamous cell carcinoma. Tumor Boil. 2018, 40.

[82] Nakagaki, T.; Tamura, M.; Kobashi, K.; Koyama, R.; Fukushima, H.; Ohashi, T.; Idogawa, M.; Ogi, K.; Hiratsuka, H.; Tokino, T.; et al. Profiling cancer-related gene mutations in oral squamous cell carcinoma from Japanese patients by targeted amplicon sequencing. Oncotarget 2017, 8, 59113–59122.

[83] Jayaprakash, C.; Varghese, V.K.; Jayaram, P.; Chakrabarty, S.; Kudva, A.; Ray, S.; Satyamoorthy, K. Relevance and actionable mutational spectrum in oral squamous cell carcinoma. J. Oral Pathol. Med. 2020, 49, 427–434.

[84] Ma, J.; Fu, Y.; Tu, Y.-Y.; Liu, Y.; Tan, Y.-R.; Ju, W.-T.; Pickering, C.R.; Myers, J.N.; Zhang, Z.-Y.; Zhong, L.-P. Mutation allele frequency threshold does not a ect prognostic analysis using next-generation sequencing in oral squamous cell carcinoma. BMC Cancer 2018, 18, 758.

[85] Er, T.K.; Wang, Y.-Y.; Chen, C.-C.; Herreros-Villanueva, M.; Liu, T.-C.; Yuan, S.-S.F. Molecular characterization of oral squamous cell carcinoma using targeted next-generation sequencing. Oral Dis. 2015, 21, 872–878.

[86] Batta, N.; Pandey, M. Mutational spectrum of tobacco associated oral squamous carcinoma and its therapeutic significance. World J. Surg. Oncol. 2019, 17, 1–12.

[87] Chen, T.-W.; Lee, C.-C.; Liu, H.; Wu, C.-S.; Pickering, C.R.; Huang, P.-J.; Wang, J.; Chang, I.Y.-F.; Yeh, Y.-M.; Chen, C.-D.; et al. APOBEC3A is an oral cancer prognostic biomarker in Taiwanese carriers of an APOBEC deletion polymorphism. Nat. Commun. 2017, 8, 465.

[88] Oikawa, Y.; Morita, K.-I.; Kayamori, K.; Tanimoto, K.; Sakamoto, K.; Katoh, H.; Ishikawa, S.; Inazawa, J.; Harada, H. Receptor tyrosine kinase amplification is predictive of distant metastasis in patients with oral squamous cell carcinoma. Cancer Sci. 2017, 108, 256–266.

[89] Van Ginkel, J.H.; De Leng, W.W.; De Bree, R.; Van Es, R.J.; Willems, S.M. Targeted sequencing reveals TP53 as a potential diagnostic biomarker in the post-treatment surveillance of head and neck cancer. Oncotarget 2016, 7, 61575–61586.

[90] Tabatabaeifar, S.; Larsen, M.J.; Larsen, S.R.; Kruse, T.A.; Thomassen, M.; Sørensen, J.A. Investigating a case of possible field cancerization in oral squamous cell carcinoma by the use of next-generation sequencing.Oral Oncol. 2017, 68, 74–80.

[91] Chen, S.-J.; Liu, H.; Liao, C.-T.; Huang, P.-J.; Huang, Y.; Hsu, A.; Tang, P.; Chang, Y.-S.; Chen, H.-C.; Yen, T.-C. Ultra-deep targeted sequencing of advanced oral squamous cell carcinoma identifies a mutation-based prognostic gene signature. Oncotarget 2015, 6, 18066–18080.

[92] Zammit, Andrew P., Rohit Sinha, Caroline L. Cooper, Christopher F. L. Perry, Ian H. Frazer, and Zewen K. Tuong. 2018. «Examining the Contribution of Smoking and HPV Towards the Etiology of Oral Cavity Squamous Cell Carcinoma Using High-Throughput Sequencing: A Prospective Observational Study.» PLoS One 13 (10): e0205406. https://doi.org/10.1371/journal.pone.0205406.

[93] Gao, Haihe, Lisha Li, Mang Xiao, Yongwei Guo, Yi Shen Lixin Cheng, and Ming Tang. 2018. «Elevated DKK1 Expression Is an Independent Unfavorable Prognostic Indicator of Survival in Head and Neck Squamous Cell Carcinoma.» Cancer Management and Research 10: 5083–89. https://doi.org/10.2147/CMAR.S177043.

[94] Hanna, Glenn J., Sook-Bin Woo, Y. Y. Li, Justine A. Barletta, Peter S. Hammerman, and Jochen H. Lorch. 2018. «Tumor PD-L1 Expression Is Associated with Improved Survival and Lower Recurrence Risk in Young Women with Oral Cavity Squamous Cell Carcinoma.» International Journal of Oral and Maxillofacial Surgery 47 (5): 568–77. https://doi.org/10.1016/j. ijom.2017.09.006

[95] Izumchenko, E.; Sun, K.; Jones, S.; Brait, M.; Agrawal, N.; Koch, W.; Mccord, C.L.; Riley, D.R.; Angiuoli, S.; Velculescu, V.E.; et al. Notch1 mutations are drivers of oral tumorigenesis. Cancer Prev. Res. 2014, 8, 277–286.

[96] Yang, CM., Wang, TH., Chen, HC. *et al.* Aberrant DNA hypermethylation-silenced SOX21-AS1 gene expression and its clinical importance in oral cancer. *Clin Epigenet* 8, 129 (2016). https://doi.org/10.1186/s13148-016-0291-5.

[97] Swati Shree Padhi, Souvick Roy, Madhabananda Kar, Arka Saha, Shomereeta Roy, Amit Adhya, Manas Baisakh, Birendranath Banerjee, Role of CDKN2A/p16 expression in the prognostication of oral squamous cell carcinoma, Oral Oncology, Volume 73, 2017, Pages 27-35, ISSN 1368-8375, https://doi.org/10.1016/j.oraloncology.2017.07.030.

[98] Evgeny Izumchenko, Kai Sun, Sian Jones, Mariana Brait, Nishant Agrawal, Wayne Koch, Christine L. McCord, David R. Riley, Samuel V. Angiuoli, Victor E. Velculescu, Wei-Wen Jiang, David Sidransky; Notch1 Mutations Are Drivers of Oral Tumorigenesis. Cancer Prev Res (Phila) 1 April 2015; 8 (4): 277–286. https://doi.org/10.1158/1940-6207.CAPR-14-0257.

[99] Maria Schwaederle, Ranajoy Chattopadhyay, Shumei Kato, Paul T. Fanta, Kimberly C. Banks, In Sil Choi, David E. Piccioni, Sadakatsu Ikeda, AmirAli Talasaz, Richard B. Lanman, Lyudmila Bazhenova, Razelle Kurzrock; Genomic Alterations in Circulating Tumor DNA from Diverse Cancer Patients Identified by Next-Generation Sequencing. Cancer Res 1 October 2017; 77 (19): 5419–5427. https://doi.org/10.1158/0008-5472.CAN-17-0885.

[100] Park, Ha Young, Joong Seob Lee, Jee Hye Wee, Jeong Wook Kang, Eun Soo Kim, Taeryool Koo, Hee Sung Hwang, Hyo Jung Kim, Ho Suk Kang, Hyun Lim, and et al. 2023. «Assessment of the Mutation Profile of Tonsillar Squamous Cell Carcinomas Using Targeted Next-Generation Sequencing» *Biomedicines* 11, no. 3: 851. https://doi.org/10.3390/biomedicines11030851.

[101] Haque, M.M., Kowtal, P. & Sarin, R. Identification, and characterization of *TP53* gene Allele Dropout in Li-Fraumeni syndrome and Oral cancer cohorts. *Sci Rep* 8, 11705 (2018). https://doi.org/10.1038/s41598-018-30238-7.

[102] Kenya Kobayashi, Seiichi Yoshimoto, Mizuo Ando, Fumihiko Matsumoto, Naoya Murakami, Go Omura, Yoshitaka Honma, Yoshifumi Matsumoto, Atsuo Ikeda, Azusa Sakai, Kohtaro Eguchi, Akiko Ito, Eigitsu Ryo, Yasushi Yatabe, Taisuke Mori, Full-coverage TP53 deep sequencing of recurrent head and neck squamous cell carcinoma facilitates prognostic assessment after recurrence, Oral Oncology, Volume 113, 2021, 105091, ISSN 1368-8375 https://doi.org/10.1016/j.oraloncology.2020.105091.

[103] Sundaresan, S., & Selvaraj, L. (2023). Role of NGS in Oral Squamous Cell Carcinoma. IntechOpen. doi: 10.5772/intechopen.108179.

[104] Samman, M., Wood, H.M., Conway, C., Stead, L., Daly, C., Chalkley, R., Berri, S., Senguven, B., Ross, L., Egan, P., Chengot, P., Ong, T.K., Pentenero, M., Gandolfo, S., Cassenti, A., Cassoni, P., Al Ajlan, A., Samkari, A., Barrett, W., MacLennan, K., High, A. and Rabbitts, P. (2015), A novel genomic signature reclassifies an oral cancer subtype. Int. J. Cancer, 137: 2364-2373. https://doi.org/10.1002/ijc.29615.

[105] Lin, Li-Han, Cheng, Hui-Wen, and Liu, Chung-Ji. 'Droplet Digital Polymerase Chain Reaction for Detection and Quantification of Cell-free DNA TP53 Target Somatic Mutations in Oral Cancer'. 1 Jan. 2022: 29 – 41.

[106] Santos, JN, Sousa Neto, ES, França, JA, et al. Next generation sequencing of oncogenes and tumor suppressor genes in odontogenic myxomas. *J Oral Pathol Med*. 2017; 46: 1036–1039. https://doi.org/10.1111/jop.12598.

[107] A. Moreira, A. Poulet, J. Masliah-Planchon, C. Lecerf, S. Vacher, L. Larbi Chérif, C. Dupain, G. Marret, E. Girard, L. Syx, C. Hoffmann, E. Jeannot, J. Klijanienko, I. Guillou, O. Mariani, A. Dubray-Vautrin, N. Badois, M. Lesnik, O. Choussy, V. Calugaru, E. Borcoman, S. Baulande, P. Legoix, B. Albaud, N. Servant, I. Bieche, C. Le Tourneau, M. Kamal, Prognostic value of tumor mutational burden in patients with oral cavity squamous cell carcinoma treated with upfront surgery, ESMO Open, Volume 6, Issue 4, 2021, 100178, ISSN 2059-7029, https://doi.org/10.1016/j.esmoop.2021.100178. (https://www.sciencedirect.com/science/article/pii/S2059702921001381).

[108] Yi, Y., Tian, Z., Ju, H., Ren, G., & Hu, J. (2017). A novel NOTCH3 mutation identified in patients with oral cancer by whole exome sequencing. International Journal of Molecular Medicine, 39, 1541-1547. https://doi.org/10.3892/ijmm.2017.2965.

[109] Tabatabaeifar, S., Kruse, T. A., Thomassen, M., Larsen, M. J., & Sørensen, J. A. (2015). *Use of next generation sequencing in oral cavity cancer: intra tumour heterogeneity and metastasis*. Abstract from DSPR Forårsmøde 2015, København, Denmark. http://dspr.dk/Content/uploads/65c8d146-195b-425e-b597-b0c2c35d1ebc.pdf

[110] Chung Ji Liu, Shu-Chun Lin, Li-Han Lin; Abstract 1234: Somatic mutations of FAT1 in oral cancer are associated with tumor progression and survival. *Cancer Res* 1 July 2018; 78 (13_Supplement): 1234. https://doi.org/10.1158/1538-7445.AM2018-1234.

[111] Hyodo, T., Kuribayashi, N., Fukumoto, C. *et al.* The mutational spectrum in whole exon of *p53* in oral squamous cell carcinoma and its clinical implications. *Sci Rep* 12, 21695 (2022). https://doi.org/10.1038/s41598-022-25744-8.

[112] Zammit, Andrew. «Aetiology of Oral Cavity Squamous Cell Carcinoma An Exploration utilising Next Generation Sequencing.» (2017).

[113] Leon J. Wils, Jos B. Poell, Arjen Brink, Ilkay Evren, Elisabeth R. Brouns, Jan G.A.M. de Visscher, Elisabeth Bloemena, Ruud H. Brakenhoff; Elucidating the Genetic Landscape of Oral Leukoplakia to Predict Malignant Transformation. Clin Cancer Res 1 February 2023; 29 (3): 602–613. https://doi.org/10.1158/1078-0432.CCR-22-2210.

[114] Avaniyapuram Kannan Murugan, Arasambattu Kannan Munirajan, Nobuo Tsuchida, Genetic deregulation of the PIK3CA oncogene in oral cancer, Cancer Letters, Volume 338, Issue 2, 2013, Pages 193-203, ISSN 0304-3835, https://doi.org/10.1016/j.canlet.2013.04.005.

[115] Keisuke Sawada, Shuji Momose, Ryutaro Kawano, Masakazu Kohda, Tarou Irié, Kenji Mishima, Takahiro Kaneko, Norio Horie, Yasushi Okazaki, Morihiro Higashi, Jun-ichi Tamaru, Immunohistochemical staining patterns of p53 predict the mutational status of TP53 in oral epithelial dysplasia, Modern Pathology, Volume 35, Issue 2, 2022, Pages 177-185, ISSN 0893-3952, https://doi.org/10.1038/s41379-021-00893-9.

[116] Sehui Kim, Chung Lee, Hyangmi Kim, Sun Och Yoon, Genetic characteristics of advanced oral tongue squamous cell carcinoma in young patients, Oral Oncology, Volume 144, 2023, 106466, ISSN 1368-8375, https://doi.org/10.1016/j.oraloncology.2023.106466.

[117] Morse, Douglas E., Walter J. Psoter, Deborah Cleveland, Donald Cohen, Mireseyed Mohit-Tabatabai, Diane L. Kosis, and Ellen Eisenberg. 2007. «Smoking and Drinking in Relation to Oral Cancer and Oral Epithelial Dysplasia.» Cancer Causes & Control 18 (9): 919–29. https://doi.org/10.1007/s10552-007-9026-4.

[118] Tannock, I.F.; Hill, R.P.; Carey, R.W. The Basic Science of Oncology. Plast. Reconstr. Surg. 1989, 83, 920.

[119] Levine, Arnold J., A. Finlay Cathy, and W. Hinds Philip. 2004. «P53 Is a Tumor Suppressor Gene.» Cell 116 (2 Suppl): S67–S69. https://doi.org/10.1016/s0092-8674 (04) 00036-4.

[120] Singh, Ragini D., R. Patel Kinjal, and S. Patel Prabhudas. 2016. «p53 Mutation Spectrum and Its Role in Prognosis of Oral Cancer Patients: A Study From Gujarat, West India.» Mutation Research 783 (January): 15–26. https://doi.org/10.1016/j.mrfmmm.2015.12.001

[121] Ragos, Vasileios, S. Mastronikolis Nicholas, Evangelos Tsiambas, Evangelia Baliou, Stylianos N. Mastronikolis, Nikolaos Tsoukalas, Eugenia E. Patsouri, and Panagiotis P. Fotiades. 2018 «P53 Mutations in Oral Cavity Carcinoma.» Journal of B.U.ON.

[122] Foulkes, William D., Tamar Y. Flanders, Pamela M. Pollock, and Nicholas K. Hayward. 1997. «The CDKN2A (P16) Gene and Human Cancer.» Molecular Medicine (Cambridge, Mass) 3 (1): 5–20. http://www.ncbi.nlm.nih.gov/pubmed/9132280.

[123] Roshan, V. G. Deepak, M. S. Sinto, Bipin T. Vargees, and S. Kannan. «Loss of CDKN2A and CDKN2B Expression Is Associated With Disease Recurrence in Oral Cancer.» Journal of Oral and Maxillofacial Pathology: JOMFP 23 (1): 82–89. https://doi.org/10.4103/jomfp.JOMFP_184_18.

[124] Riese, U., R. Dahse, W. Fiedler, C. Theuer, S. Koscielny, G. Ernst, E. Beleites, U. Claussen, and F. von Eggeling. 1999. «Tumor Suppressor Gene P16 (CDKN2A) Mutation Status and Promoter Inactivation in Head and Neck Cancer.» International Journal of Molecular Medicine 4 (1): 61–65. https://doi.org/10.3892/ijmm.4.1.61.

[125] Andrew P. Read, Susan Holland, Stephan Porter, et al. 1999. «DNA Studies Underestimate the Major Role of CDKN2A Inactivation in Oral and Oropharyngeal Squamous Cell Carcinomas.» Genes, Chromosomes & Cancer 25 (1): 16–25. http://www.ncbi.nlm.nih.gov/pubmed/10221335.

[126] Lim, Annette M., Hongdo Do, Richard J. Young, Stephen Q. Wong, Christopher Angel, Marnie Collins, Elena A. Takano, et al. 2014. «Differential Mechanisms of CDKN2A (P16) Alteration in Oral Tongue Squamous Cell Carcinomas and Correlation With Patient Outcome.» International Journal of Cancer 135 (4): 887–95. https://doi.org/10.1002/ijc.28727.

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